

Mechanism of Teratogenic Action of Hypoglycin-A

Hypoglycin-A¹ is a toxic non-proteinogenic amino acid of considerable biochemical interest^{2,3}. It has been suggested that the toxicity of hypoglycin-A is due to the formation of a degradation product, methylenecyclopropane acetate, which inhibits the oxidation of long-chain fatty acids⁴. ENTMAN and BRESSLER⁵ reported that L-carnitine, which promotes the intramitochondrial oxidation of long-chain fatty acids^{6,7}, antagonized the hypoglycaemic effects of hypoglycin-A. Hypoglycin-A inhibited the growth of sarcoma-180 cells in mice, but a simultaneous injection of leucine, a structural analogue of hypoglycin-A, prevented this inhibition of tumour growth⁸.

In the pregnant rat, hypoglycin-A is highly teratogenic⁹. The present communication describes the effects on embryonic development, in pregnant rats, of administering hypoglycin-A simultaneously with riboflavin phosphate, L-leucine, or DL-carnitine.

Materials and methods. The methods used in the present studies were previously described⁹. Pregnant Wistar-derived rats (160–180 g) were divided into 4 groups. L-leucine (15 mg/kg) and hypoglycin-A (30 mg/kg) were administered to animals of Group I. Group II received riboflavin phosphate (3 mg/kg) and hypoglycin-A (30 mg/kg), Group III DL-carnitine (200 mg/kg) and hypoglycin-A (30 mg/kg), and Group IV animals received only hypoglycin-A (30 mg/kg) and served as a control. Hypoglycin-A, L-leucine, riboflavin phosphate, and DL-carnitine were dissolved in isotonic saline, the pH being adjusted to 7.4 with phosphate buffer, and administered i.p. from the first through the sixth day of gestation.

Results and discussion. The results are summarized in the Table. Simultaneous administration of leucine and hypoglycin-A to the pregnant rats did not counteract the teratogenic effect of hypoglycin-A, but resulted in a significant increase in the incidence of foetal resorption, compared with animals treated with only hypoglycin-A

($P < 0.01$). The simultaneous injection of carnitine did not prevent the teratogenic effects of hypoglycin-A, but the combined treatment caused marked embryotoxic and teratogenic effects which showed no significant difference compared with the hypoglycin-A treated animals. The incidence of foetal abnormalities was significantly reduced ($P < 0.001$) in the pregnant animals treated with both hypoglycin-A and riboflavin phosphate compared with the controls. However, the number of foetal resorptions was not significantly affected.

Inhibition of long-chain fatty acid oxidation is considered to be the primary effect of hypoglycin-A on intermediary metabolism²⁻⁴. Accumulation of fatty acids, uncoupling of oxidative phosphorylation, and the resulting block in the availability of energy for ATP synthesis may account for the teratogenicity of hypoglycin-A, since ATP is essential for normal embryonic growth and differentiation¹⁰. Reversal of hypoglycin-A induced teratogenic effects by riboflavin phosphate lends support to the suggestion of VON HOLT et al.⁴ that the actual site of action of hypoglycin-A is inhibition of the acyl dehydrogenase flavin-dependent-oxidation reaction.

Zusammenfassung. Simultane Verabreichung von Riboflavin und Hypoglycin-A reduzierte das Auftreten von foetalen Missbildungen in Ratten. Dagegen haben Carnitin und Leucin keinen Einfluss auf die teratogene Wirkung des Hypoglycins.

T. V. N. PERSAUD¹¹

Department of Anatomy, University of the West Indies, Mona, Kingston 7 (Jamaica), 9 July 1970.

Influence of leucine, riboflavin phosphate and carnitine on hypoglycin-A induced teratogenicity

Groups (No. of animals)	Treat- ment	Total implan- tations	Resorptions	Malformed foetuses
I (6)	H + L	52	17 (32.7%)	35 (100%)
II (6)	H + R	51	3 (5.9%)	21 (43.7%)
III (6)	H + C	48	3 (6.3%)	36 (80%)
IV (6)	H	53	4 (7.5%)	40 (81.6%)

H, hypoglycin-A; L, leucine; C, carnitine; R, riboflavin phosphate.

¹ C. H. HASSALL, K. REYLE and P. FENG, *Nature*, Lond. 173, 356 (1954).

² R. BRESSLER, C. CORREDOR and K. BRENDL, *Pharmac. Rev.* 21, 105 (1969).

³ H. S. A. SHERRATT, *Br. med. Bull.* 25, 250 (1969).

⁴ C. VON HOLT, M. VON HOLT and H. BÖHM, *Biochim. biophys. Acta* 125, 11 (1966).

⁵ M. ENTMAN and R. BRESSLER, *Molec. Pharmac.* 3, 333 (1967).

⁶ J. BREMER, *J. biol. Chem.* 237, 3628 (1962).

⁷ M. KLINGENBERG and C. BODE, in *Recent Researches on Carnitine* (Ed. G. WOLF; M.I.T. Press, Cambridge, Mass. 1965), p. 87.

⁸ R. GASKIN and T. V. N. PERSAUD, *W. Indian med. J.* 16, 198 (1967).

⁹ T. V. N. PERSAUD, *W. Indian med. J.* 16, 193 (1967).

¹⁰ E. J. BOELL, in *Analysis of Development* (Eds. B. H. WILLIER, P. A. WEISS and V. HAMBURGER; W. B. Saunders Company, Philadelphia and London 1955), p. 520.

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Uptake of Aflatoxin B₁ by Plastic Materials¹

Aflatoxins are a group of mycotoxins that have gained increasing interest as food contaminants during the past 8 years. Their hepatotoxic properties have led to extensive research on the chemical and physical characteristics of aflatoxins in an effort to determine their mode of action². Since the carcinogenic properties of aflatoxins

in animals, including different species of monkeys, have been well documented³⁻⁶, these mycotoxins are suspected as being responsible for the high incidence of liver cancer in certain areas⁷.

An investigation on aflatoxin B₁ was initiated in our laboratory: it included metabolism in perfused rat liver